

a2 Identification of the SIM1 and ARNT DNA fragments from the yeast two-hybrid plasmid carried by the clone 06L22 by duplex PCR. Plasmid DNA was isolated from a liquid culture of the clone 06L22 by a QiaPrep (Hilden) procedure and the inserts contained within the plasmids were amplified by PCR using the primer pairs, 5'-TCG TAG ATC TTC GTC AGC AG-3' (SEQ ID No. 4) & 5'-GGA ATT AGC TTG GCT GCA GC-3' (SEQ ID No. 5) for the plasmid pBTM117c and 5'-CGA TGA TGA AGA TAC CCC AC-3' (SEQ ID No. 6) & 5'-GCA CAG TTG AAG TGA ACT TGC-3' (SEQ ID No. 7) for pGAD427. Lane 1

Please replace the fifth full paragraph on page 51 with:

a3 For the construction of pGAD427 a 1.2 kb *Dde* I fragment containing the *aphA* gene was isolated from pFG101u (Pansegrau et al., 1987) and was subcloned into the *Pvu* I site of the pGAD426 using the oligonucleotide adapters 5'- GTCGCGATC-3' and 5'-TAAGATCGCGACAT-3' (SEQ ID No. 8). The plasmid pGAD426 was generated by insertion of a 1.2 kb *Eco* RV *CYH2* gene fragment, which was isolated from the pAS2-1 (Clonetech) into the *Pvu* II site of pGAD425 (Han and Collicelli, 1995).

Please replace the fourth full paragraph on page 90 with:

a4 Plasmids carrying the mammalian readout systems named pG5E1bEGFPneo, pG5E1bEGFP_{hyg} or pG5E1bEGFP_{pur} are used. These plasmids contain the TATA element of the adenoviral E1b gene and five tandem copies of the GAL4 responsive element UAS_G (5' CGGAGTACTGTCC TGCG 3' (SEQ ID No. 9)) (Sadowski, I., Ma, J., Treizenberg, S. and Ptashne, M. (1988), Nature 335: 559-560) positioned immediately upstream of the coding sequence for the enhanced green fluorescent protein (EGFP; Yang, T.T., Cheng, L. and Kain, S.R. (1996), Nucl. Acids Res. 24 (22): 4592-4593). These reporter plasmids are generated by replacing the coding sequence for CAT in G5E1bCAT (Dang, C.V., Barrett, J., Villa-Garcia, M., Resar, L.M.S., Kato, G.J. and Fearon, E.R. (1991), Mol. Cell. Biol. 11: 954-962) by the EGFP coding sequence and introducing either a neomycin, hygromycin or puromycin resistance marker gene (neo^r, hyg^r or pur^r) using standard subcloning procedures.

Please replace the first full paragraph on page 91 with:

a5 The plasmids pVP-Nconeo and pVP-Ncohyg are derived from pVP-Nco vector (Tsan, J., Wang, Z., Jin, Y., Hwang, L., Bash, R.O., Baer, R. The Yeast Two-Hybrid System, edn 1. Edited by Bartel, P.L., Fields, S. New York: Oxford University Press (1997): 217-232) by insertion of either a neo^r or hyg^r marker gene using standard subcloning procedures. pVP-Nco in turn is an improved version of the pNLVP16 plasmid, which was constructed for the expression of herpes simplex virus protein VP16-fusion proteins in mammalian cells (Dang, C.V., Barrett, J., Villa-Garcia, M., Resar, L.M.S., Kato, G.J. and Fearon, E.R. (1991), Mol. Cell. Biol. 11: 954-962). A polylinker sequence is preceded by an artificial reading frame including the eleven amino-terminal residues of Gal4p (MKLLSSIEQAC (SEQ ID No. 10)), a nuclear localization signal from the SV40 large T antigen (PKKKRKVD (SEQ ID No. 11)) and the acidic transactivation domain (amino acids 411-456) of the VP16 protein. Alternatively, the hybrid reading frames comprising Gal4 (1-147) and individual sequences of a cDNA library are subcloned into pLXSN or any other similar retroviral vector to allow packaging cell line-aided infection of target cells.

Please replace the table on page 101 with:

a6

Oligonucleotide	Sequence (5'-3')
a sense	TCGAGTCGACGCGGCCGCTAA (SEQ ID No. 12)
A antisense	GGCCTTAGCGGCCGCGTCGAC (SEQ ID No. 13)
b sense	TCGAGGTCGACGCGGCCGCGAGTAA (SEQ ID No. 14)
B antisense	GGCCTTACTGCGGCCGCGTCGACC (SEQ ID No. 15)
c sense	TCGAGAGTCGACGCGGCCGCTTAA (SEQ ID No. 16)
c antisense	GGCCTTAAGCGGCCGCGTCGACTC (SEQ ID No. 17)

Please enter the sequence listing filed herewith.

The replacement paragraphs presented above incorporate changes as indicated by the marked-up versions below.

Polylinkers used within the multiple cloning site to provide expression of the genetic fragment in one of the three reading frames. (SEQ ID Nos. 1-3)

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For the construction of pGAD427 a 1.2 kb *Dde* I fragment containing the *aphA* gene was isolated from pFG101u (Pansegrau et al., 1987) and was subcloned into the *Pvu* I site of the pGAD426 using the oligonucleotide adapters 5'- GTCGCGATC-3' and 5'-TAAGATCGCGACAT-3' (SEQ ID No. 8). The plasmid pGAD426 was generated by insertion of a 1.2 kb *Eco* RV *CYH2* gene fragment, which was isolated from the pAS2-1 (Clonotech) into the *Pvu* II site of pGAD425 (Han and Collicelli, 1995).

Plasmids carrying the mammalian readout systems named pG5E1bEGFPneo, pG5E1bEGFP^{hyg} or pG5E1bEGFP^{pur} are used. These plasmids contain the TATA element of the adenoviral E1b gene and five tandem copies of the GAL4 responsive element UAS_G (5' CCGAGTACTGTCC TGCG 3' (SEQ ID No. 9)) (Sadowski, I., Ma, J., Treizenberg, S. and Ptashne, M. (1988), Nature 335: 559-560) positioned immediately upstream of the coding sequence for the enhanced green fluorescent protein (EGFP; Yang, T.T., Cheng, L. and Kain, S.R. (1996), Nucl. Acids Res. 24 (22): 4592-4593). These reporter plasmids are generated by replacing the coding sequence for CAT in G5E1bCAT (Dang, C.V., Barrett, J., Villa-Garcia, M., Resar, L.M.S., Kato, G.J. and Fearon, E.R. (1991), Mol. Cell. Biol. 11: 954-962) by the EGFP coding sequence and introducing either a neomycin, hygromycin or puromycin resistance marker gene (neo^r, hyg^r or pur^r) using standard subcloning procedures.